

# *Helicobacter pylori* and its role in the pathogenesis of follicular gastritis: an overview

## *Helicobacter pylori* y su rol en la patogénesis de la gastritis folicular: una visión general

Yeison Carlosama-Rosero<sup>1,2,3</sup> , Gonzalo Latorre<sup>4</sup> , Arnoldo Riquelme<sup>4,5</sup> , José Darío Portillo-Miño<sup>6,7</sup> 

<sup>1</sup> Interdisciplinary Research Group on Health-Disease (GIISE), Universidad Cooperativa de Colombia. Campus Pasto. Nariño. Colombia.

<sup>2</sup> Department of Pathology, Universidad Cooperativa de Colombia, Campus Pasto. Nariño. Colombia.

<sup>3</sup> Health Sciences Doctoral Program, Universidad Autónoma de Manizales, Manizales. Colombia.

<sup>4</sup> Department of Gastroenterology, Faculty of Medicine, Pontificia Universidad Católica de Chile, Santiago. Chile.

<sup>5</sup> Centro para la Prevención y Control del Cáncer (CECAN), Santiago. Chile.

<sup>6</sup> Infectious Diseases and Cancer Research Group, Centro de Investigaciones Clínicas, Fundación Hospital San Pedro, Pasto, Nariño, Colombia.

<sup>7</sup> Colombian Research Group on *Helicobacter pylori*, Bogotá D.C., Colombia.

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### Correspondence:

José Darío Portillo Miño,  
Street 16 Av. 43, Neighborhood San Pedro, San Juan de Pasto.  
520003. Nariño. Colombia  
Telephone: PBX (602) 7 336000 Ext 4250  
E-mail: cic.investigaciones@hospital-sanpedro.org

### ABSTRACT

The role of *Helicobacter pylori* in the pathogenesis of peptic ulcers and gastric adenocarcinoma is widely known; however, it is not entirely understood how bacterial infection is closely related to the genesis of follicular gastritis and some types of gastric lymphoma. Diagnosing and pathogenic mechanisms follicular gastritis remain challenging. Therefore, this article aims to examine the role of *H. pylori* in the development of follicular gastritis. In addition, it emphasizes the clinical and histopathological fundamentals. A broader overview of follicular gastritis is presented, and implementing preventive strategies, such as *H. pylori* eradication remains the standard treatment.

**Keywords:** *Helicobacter pylori*; *Pseudolymphoma*; *Lymphoid Hyperplasia*, *Follicles* (source: MeSH NLM).

### RESUMEN

El papel de *Helicobacter pylori* en la patogénesis de las úlceras pépticas y el adenocarcinoma gástrico es ampliamente conocido; sin embargo, no se entiende por completo cómo la infección bacteriana está estrechamente relacionada con la génesis de la gastritis folicular y algunos tipos de linfoma gástrico. El diagnóstico y los mecanismos patogénicos de la gastritis folicular siguen siendo un desafío. Por lo tanto, este artículo tiene como objetivo examinar el papel de *H. pylori* en el desarrollo de la gastritis folicular. Además, enfatiza los fundamentos clínicos e histopatológicos. Se presenta una visión general más amplia de la gastritis folicular y la implementación de estrategias preventivas, como la erradicación de *H. pylori*, que sigue siendo el tratamiento estándar.

**Palabras clave:** *Helicobacter pylori*; *Pseudolinfoma*; *Hiperplasia Linfoide*, *Foliculos* (fuente: DeCS Bireme).

### INTRODUCTION

Follicular gastritis (FG) consists of a particular chronic persistent inflammatory process, which is preceded by *Helicobacter pylori* (*H. pylori*) infection<sup>(1,2)</sup>. These bacteria are estimated to have infected more than 50% of the global population<sup>(3-6)</sup>. Likewise, it has been proposed that the acquisition and colonization of *H. pylori* begin in childhood and are established as a chronic infection. This triggers perpetual and progressive gastric inflammation, leading to clinical complications in about 1-10% of patients<sup>(7,8)</sup>. In this sense, *H. pylori* has been associated with diseases such as peptic ulcer, atrophic gastritis, intestinal gastritis, gastric cancer, and mucosa-associated lymphoid tissue lymphoma (MALT)<sup>(1,8)</sup>. In addition, the International Agency for Research on Cancer<sup>(9)</sup> of the World Health Organization (WHO) has classified it as a type I carcinogen<sup>(10)</sup>. It is also worth mentioning that *H. pylori* infection has been included as an infectious disease in the update of the International Classification of Diseases (ICD-11)<sup>(11)</sup>. Therefore, all infected patients must receive adequate treatment, generating a paradigm shift since treatment is no longer only reserved for patients with clinical manifestations of infection<sup>(12)</sup>.

Although some studies associate *H. pylori* infection with FG<sup>(13-19)</sup>, a high prevalence of gastric MALT lymphoma has also been observed in patients without

*H. pylori* infection<sup>(20)</sup>; Therefore, its role in the FG progression to MALT lymphoma has not yet been completely clarified. Given this scenario, an enormous interest and research on *H. pylori* infection has been aroused, primarily motivated by the need to reexamine this problem and elucidate the pathogenic mechanisms involved, the multiple conditions associated with the bacteria, virulence factors, and the heterogeneity of the disease. Therefore, it is pertinent to carry out this Article Review that aims to expose the critical role that *H. pylori* play in the development of FG, the histopathological findings, etiopathogenesis, and diagnostic methods to provide a comprehensive overview of FG. Therefore, we suggest implementing strategies such as monitoring, and *H. pylori* eradication for the treatment of FG.

### Definition

FG is a particular type of chronic gastritis characterized by the presence of a mononuclear inflammatory infiltrate and the formation of lymphoid follicles with a germinal center<sup>(21,22)</sup>. Although several studies have made a closer approximation of the macroscopic findings of FG, the microscopic definition is not well-established<sup>(23)</sup>. This limitation has led to the lack of an acceptable definition of histopathological findings. Therefore, the use of confusing definitions and the absence of standardization between studies has made it difficult to establish the cause-effect relationship and delay the diagnosis<sup>(23)</sup>. At least, for most relevant studies, FG is characterized by follicular lymphoid hyperplasia of lymphoid nodules with intraepithelial lymphocytosis<sup>(23)</sup>. In Eastern countries, the definition is more rigorous, which includes the identification of at least two secondary lymphoid follicles in an area of 1 cm of gastric mucosa in the lesser curvature of the antrum<sup>(14)</sup>. This definition is compatible with the premise that the antral mucosa is the most common location where lymphoid follicles are found. It is necessary to specify that the presence of the germinal center differentiates FG from lymphoid aggregates. In addition, there are multiple descriptions in the literature of the disease, such as follicular lymphoid hyperplasia, pseudolymphoma, nodular gastritis, nodular antritis, and lymphofollicular gastritis.

### Epidemiology

The prevalence of *H. pylori* infection varies worldwide; it ranges between 19% and 88%, mainly influenced by socioeconomic conditions, geographical location, age, and hygiene conditions<sup>(3,24,25)</sup>. Regarding the prevalence of FG, there is dissimilar data across different countries. Miyamoto *et al.*<sup>(15)</sup>, reported in Japan, a prevalence of 0.2%. Data provided by Greek and French researchers show a prevalence of 13% and 14%, respectively<sup>(26)</sup>. Unlike Korea, a study reveals a prevalence of 51%<sup>(27)</sup>. In similar studies, prevalences have been shown to range between 27% and 80%<sup>(28-31)</sup>. While other authors have estimated a prevalence of 0.7%-2.9%<sup>(15,32-34)</sup>.

In Colombia, a study carried out by Martínez-Henao *et al.* determined a prevalence of 8.4%<sup>(35)</sup>. In the recent study by Melo-Peñalosa *et al.*<sup>(36)</sup>, a FG prevalence rate of

34% was reported in patients with *H. pylori* infection and 10% without *H. pylori* infection. These heterogeneous data are likely explained by the histopathological definition, the number of gastric biopsies taken in the studies, and the site from which the was collected<sup>(31)</sup>. Therefore, an underdiagnosis of FG has been related to an insufficient number or inappropriate site of gastric samples, given that lymphoid follicles are greater by up to 44% when samples are taken from the antral mucosa compared to the findings from the body mucosa<sup>(29,37)</sup>. Additionally, the definition of FG in some studies does not discriminate between lymphoid aggregates and lymphoid follicles with a germinal center, the latter being the histopathological condition that defines the diagnosis of FG<sup>(21-23)</sup>.

### Risk factors

In the risk factors for FG, a higher frequency has been determined in pediatric patients with persistent symptoms of dyspepsia, while in females; apparently, there is a particular risk for the development of this condition<sup>(17,18,22,37-42)</sup>. The higher incidence of functional dyspepsia in females suggests a hormonal influence on the development of the condition despite the prevalent *H. pylori* infection in both genders. However, this pattern has not been consistently observed in Colombia<sup>(35)</sup>. Additionally, age and sex are not the sole predisposing factors for the development of FG. This condition has been observed more frequently in patients with other states related to *H. pylori* infection, such as gastric ulcers. Furthermore, it has been suggested that the host's immune factors may contribute to the development of FG. A study conducted by Mansilla *et al.*<sup>(43)</sup>, demonstrated when matching patients with FG by age and sex, no differences were observed in the OLGA staging. However, patients with FG had a higher bacterial load in the gastric mucosa. The role of *H. pylori* infection and the development of FG is well-established<sup>(1,16,18,44)</sup>. It has been proposed that chronic inflammation related to chronic superficial gastritis associated with *H. pylori* infection leads to FG<sup>(44,45)</sup>.

Another part discusses the foundation histopathological observation that supports the central role of *H. pylori* infection in the pathogenesis of MALT lymphoma. In this sense, it has been postulated that it is the induction of gastric lymphoid follicles, representing the first stage in cancer progression in expanding lymphoid hyperplasia<sup>(44,46)</sup>. Additionally, in a systematic review conducted by Asenjo *et al.*<sup>(47)</sup>, they reported an overall prevalence of *H. pylori* infection among patients with gastric MALT lymphoma of 75% and a contrast of 60% among patients with diffuse large B-cell lymphoma (DLBCL). Although the prevalence of *H. pylori* infection among MALT lymphoma patients depends mainly on the diagnostic methods used, histological grade, and tumor invasion<sup>(47)</sup>. These studies suggest that *H. pylori* infection is essential in the pathogenesis of early lymphoma. Furthermore, before the progression to MALT lymphoma, B cell clonality can be identified in chronic gastritis before the development of overt lymphoma<sup>(39)</sup>. However, not current evidence that supporting FG and MALT lymphoma.

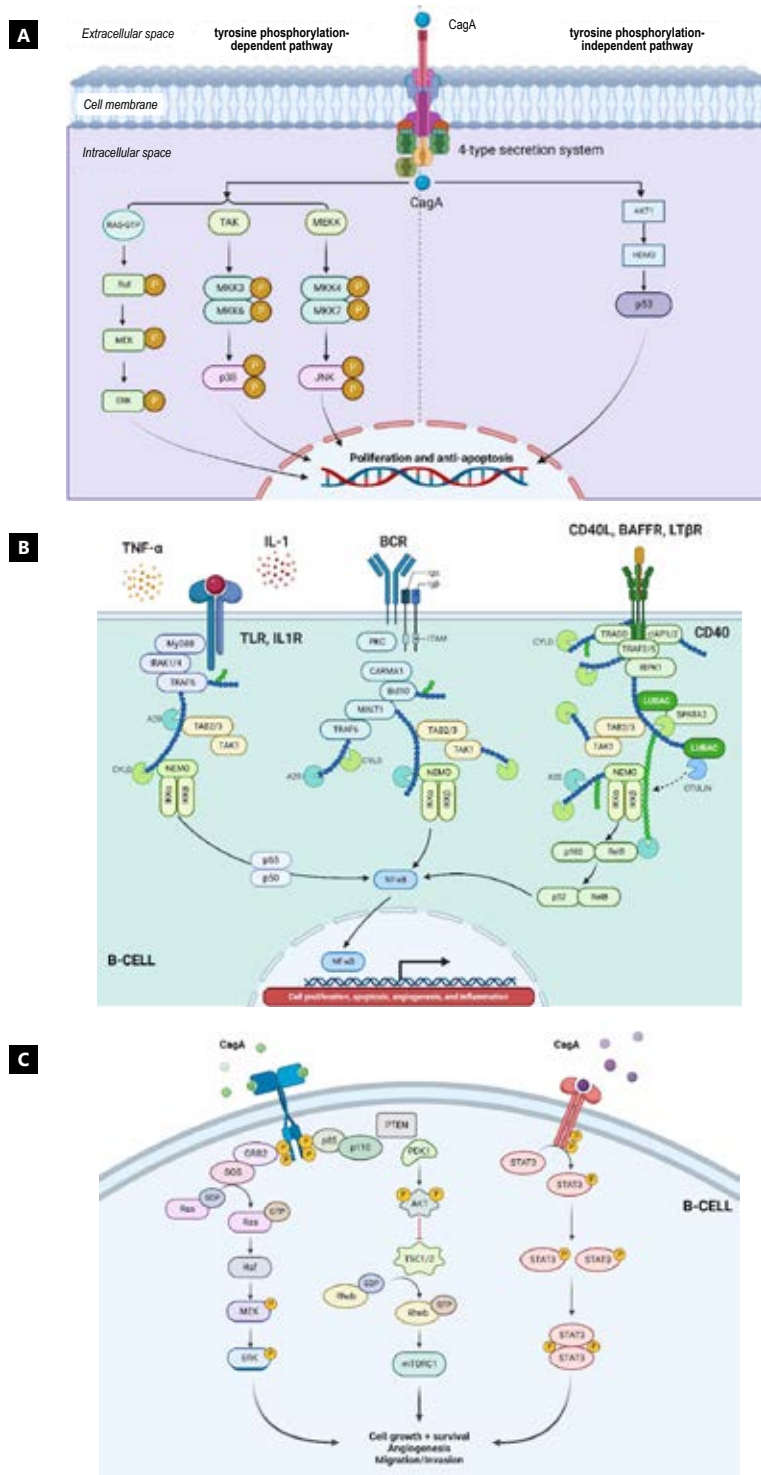
## Etiopathogenesis

*H. pylori* infection is the most common chronic infection worldwide and the leading cause of gastroduodenal disease, including chronic gastritis, peptic ulcer, gastric carcinoma, and MALT lymphoma<sup>(2,7,8,48-51)</sup>. Furthermore, a possible association between chronic autoimmune atrophic gastritis and FG has been proposed. This is due to the infiltration and destruction of the gastric mucosa by cross-reactive cytotoxic-specific T cells targeting epitopes of *H. pylori* bacteria<sup>(49)</sup>. Although the bacteria can colonize all stomach regions, the highest bacterial density has been seen in the antrum and cardia<sup>(52)</sup>, probably because in these anatomical locations, the production of hydrochloric acid is decreased<sup>(52-54)</sup>. Understanding bacterial density is essential as it has been associated with the severity of chronic stomach inflammation<sup>(55,56)</sup>. However, in the study conducted by Mansilla *et al.*<sup>(43)</sup> demonstrated increased bacterial load without a concomitant rise in mucosal inflammatory cytokine responses in *H. pylori* infection with FG. These findings suggest that the immune response is variable or an additional mechanism of *H. pylori*'s active immune evasion response. Thus, further research is necessary in this context. Aside is considered to be part of the immune response to *H. pylori* infection, which is mediated by the virulence factor cytotoxin-associated gene A (CagA)<sup>(45)</sup>. Thus, CagA is linked to significant mucosal inflammation, severe atrophic gastritis, and the development of GC<sup>(50,57,58)</sup> (see Figure 1). It has been proposed that CagA is essential in epithelial tumorigenesis since it is phosphorylated by intracellular proteins, particularly by the protein tyrosine phosphatase-2 (SHP2) that contains the Src-2 and Abl homologous domain, and induces phenotypic plasticity and oncogenesis through the phosphorylation of SHP2, MDM2, p53, NF- $\kappa$ B, ERK and the constitutive activation of the canonical phosphatidylinositol 3 kinase/AKT/mTOR pathway, considered essential in cell apoptosis and proliferation<sup>(50,59,60)</sup>. Likewise, CagA is a highly immunogenic protein that stimulates the production of interleukin-8 (IL-8) and leads to the infiltration of neutrophils in the inflammatory area, the production of free radicals, and DNA damage<sup>(59,61,62)</sup>, increasing the risk of gastric lymphomagenesis<sup>(63)</sup>. Indeed, *in vitro* experiments have shown that the immune response in extranodal marginal zone B cell lymphoma is mainly produced by T cell-mediated immunity<sup>(64)</sup>. Studies have shown that CagA antigen can translocate to B cell after destruction of the gastric mucosa during chronic gastritis<sup>(65,66)</sup>. In B cells, this antigen prevents apoptosis through extracellular signal-regulated kinase, leading to the proliferation and immortalization of B cells<sup>(65-67)</sup>. CagA antigen can induce proinflammatory responses, such as neutrophil infiltration, reactive oxygen species (ROS) production, and polyclonal B cell activation in the gastric lumen, causing gastric damage and genetic instability<sup>(59,68,69)</sup>.

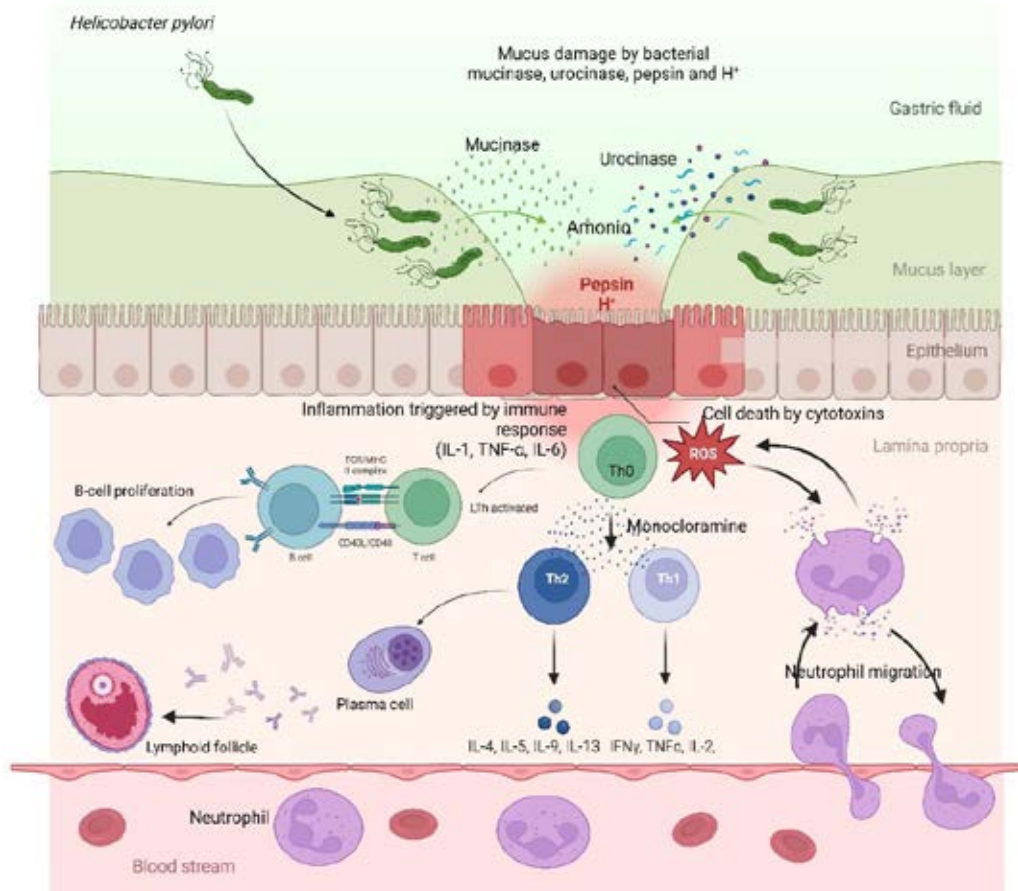
On the other hand, the bacteria's spiral shape allows it to be mobilized through the viscous gastric mucus using flagella and to avoid peristaltic sweep by attaching to the mucin through a hook-shaped protein, adhesins (BabA, SabA, OipA), and Sialyl-Lewis X antigens<sup>(70)</sup>. The

bacteria to evade the acidic microenvironment, through the enzyme urease, establish an environment formed by a cloud of ammonium plus carbon dioxide, increasing the pH<sup>(2,50,70,71)</sup>, which temporarily improves the hostile acidophilic microenvironment<sup>(52,72)</sup>. Monochloramine is an intermediate metabolite result of the combination of ammonium release of the bacterium with chloro ion of the polymorphonuclear (PMN) cells, whose harmful effects include mitochondrial damage, increased gastrin secretion, inhibition of epidermal growth factor, stasis, and alterations in microcirculation<sup>(73-77)</sup>. The cytotoxic effect of monochloramine produces the release of cytokines, which are responsible for differentiating Th0 lymphocytes into Th1 and Th2 lymphocytes. Th1 lymphocytes produce IL-2 and gamma interferon (IFN- $\gamma$ ), promoting cellular immunity, while Th2 lymphocytes produce IL-4 and IL-5, which are involved in the activation of B cells, which allows differentiation into plasma cells and production of immunoglobulins; especially, IgG and IgM antibody<sup>(78)</sup>. It has been shown that MALT lymphomas are infiltrated by Th2-polarized T lymphocytes, which favor tumor proliferation and are intensified by intratumoral CD4+ T cells<sup>(79)</sup>. Many of these CD4+ T cells are suppressors of CD25+ lymphocytes, FOXP3+ T cells, and regulatory T cells (Treg) recruited by tumor B cells. A high level of FOXP3+ expression in cells confers a better response to therapy for *H. pylori* eradication<sup>(52,79-81)</sup>. It has been shown that the immune response of patients is variable, and intricate Th1 and Th2 polarization profiles are found, which explains the heterogeneous histopathological findings in patients with *H. pylori* infection<sup>(52,82)</sup>. The result of these processes benefits the recruitment of multiple cell types, such as inflammatory cells, T cells, B cells, and plasmacytes, the formation of germinal centers, and the production of antibodies and numerous expression factors and cytokines. This evokes the extraordinary orchestration of the immune system in processes that include tumor growth, survival, progression, invasion, and metastasis<sup>(52,79-82)</sup> (see Figure 2).

Unlike Lipopolysaccharide (LPS) from other species, LPS of *H. pylori* is recognized by Toll-like receptors (TLR) such as TLR-2 instead of TLR-4<sup>(71)</sup>. Normal lymphocyte function depends on regulating the transcriptional activity of nuclear factor kappa  $\beta$  (NF- $\kappa$ B). The bacteria interacts with epithelial cells through TLR, which activates the NF- $\kappa$ B pathway, altering the regulation of signaling pathways and contributing to lymphomagenesis<sup>(83,84)</sup>, inducing release of interleukin-8 (IL-8) and stimulating the expression of intercellular adhesion molecules ICAM-1<sup>(85)</sup>. The NF- $\kappa$ B pathway is a primary transcription factor usually sequestered in the cytoplasm. It is a point of convergence of several signaling pathways activated at cell surface receptors, including the BCR, which leads to transformations in the expression of genes that modify the immune response, cell survival, proliferation, and apoptosis<sup>(84,86)</sup>. On the other hand, gastric epithelial cells express high levels of HLA-DR during chronic *H. pylori* infection, with the recruitment of T cells that express CD40 ligand (CD40L) molecules. B cells are stimulated by the CD40L-CD40 interaction associated



**Figure 1.** Signaling pathways associated with FG. **A)** The direct action of CagA on the tyrosine-kinase dependent and independent pathways. It is observed how, through the type-4 secretion system, the bacteria introduce CagA to the cell cytoplasm, which can activate tyrosine kinase phosphorylation-dependent pathways such as RAF, TAK, MEKK, and the phosphorylation-independent pathways such as AKT1 that regulates HDM2, responsible for modulating p53 called “the guardian of the genome,” which has an impact on proliferation and anti-apoptosis. **B)** The canonical NF-κB pathway is activated by cytotoxin associated with gene A (CagA) through IL-8 and its receptor, which regulate it downstream, whereas the non-canonical pathway is activated more strictly by a set of molecules known as “the members of the TNF superfamily” such as B-cell activating factor (BAFF), lymphotoxin-β and CD40 ligand (CD40L), who may possibly be responsible for the more significant activity of the non-canonical NF-κB pathway due to its role in the interaction between T cells and B cells <sup>(16)</sup>. **C)** The canonical phosphatidylinositol 3-kinase/AKT/mTOR (PI3K/AKT/mTOR) pathway is activated by the direct action of the virulence factor (CagA) of *H. pylori*, as well as by the cytokines and chemokines of the persistent chronic inflammatory response. The PI3K/AKT/mTOR pathway is essential in cell proliferation and apoptosis. Source: The authors.



**Figure 2.** The mechanisms of chronic inflammatory response by which *H. pylori* induces FG. It is observed how the bacteria, through mucinase, urokinase, pepsin, and H<sup>+</sup>, generate a decrease in the protective layer of mucus and how urokinase forms an ammonium cloud to improve the hostile environment of the bacteria. This, in turn, produces injury and cell death of the gastric epithelium, triggering chronic inflammation and a perpetual immune response with the migration of neutrophils that release proinflammatory cytokines (IL-1, TNF-α, IL-6) that activate LT. In turn, they express CD40L and TCR on their surface to interact with MHC and CD40 receptors, causing sustained and uncontrolled proliferation of BL. On the other hand, Th0, through monochloramine, is polarized into Th1, which releases molecules (IL-2, IFN-γ, TNF-α) and Th2 (IL-4, IL-5). Polarization to a Th2-type immune response allows the activation of LB and differentiates into plasma cells that produce immunoglobulins, leading to the formation of essential lymphoid follicles in FG. Source: The authors.

with the action of various cytokines and chemokines<sup>(87,88)</sup>. Furthermore, elevated expression of the APRIL ligand, a cytokine that is essential for sustained B cell proliferation, has been observed<sup>(89)</sup>, which is produced by macrophages through the induction of *H. pylori* and *H. pylori*-specific T cells<sup>(90)</sup>. Likewise, APRIL is produced by eosinophils, B cells located near lymphoid infiltrates, and tumor cells, which suggests the protumorigenic potential of APRIL<sup>(91)</sup>.

On the other hand, *Gastrokinase gene (GKN1)*, which is a tumor suppressor gene that inhibits inflammation<sup>(92)</sup>, it has been proven that its transcript and protein are decreased in the mucosa of patients with FG with *H. pylori* infection<sup>(93,94)</sup>. As mentioned above, the hormonal factor may participate in the evolution of the disease, which acts exclusively on a Th2-type response; therefore, a significant reduction in inflammatory activity during pregnancy has been demonstrated<sup>(95)</sup>. This phenomenon would explain,

to some extent, the greater prevalence of FG in the female population.

The *H. pylori* bacteria constitute the main inducing factor of an immune response that stimulates the proliferation of lymphoid tissue in the lamina propria, especially MALT, whose function is to protect the surface of the gastrointestinal tract and other exposed mucosa to the external environment. It is currently under discussion whether the activation of B cells with monoclonal proliferation results from an autoimmune response or is an intermediary of persistent stimulation by *H. pylori* antigens. In this order of ideas, it has been shown that MALT lymphoma B cells express polyreactive surface immunoglobulins (BCR) such as IgG and direct stimulation by alloantigens and autoantigens recognized by surface antibodies that lead to the proliferation of tumor cells. After this oligoclonal expansion, a dominant clonal proliferation can be exposed to the surface through

selective pressure<sup>(16,44,87)</sup>. This postulate suggests a possible explanation for the high regression rates in patients with FG and even cases of MALT lymphoma when treatment is instituted to eradicate *H. pylori* infection. Besides, the development of monoclonal MIB-1 antibodies, a reactive epitope of the nuclear antigen ki67 involved in the proliferation of epithelial cells, has been considered an indicator of cell proliferation in biopsy<sup>(96)</sup>.

A hypothesis has emerged regarding the relationship between bacterial genotypes and certain diseases. The genotypes CagA+ and vacA s1m1 are the most virulent strains associated with peptic ulcers, GC, and severe inflammation<sup>(97)</sup>, while the s2m2 strain generates limited inflammation. In this order of ideas, CagA has been frequently detected in MALT lymphomas, and the response to *H. pylori* eradication is faster in patients who express CagA<sup>(98)</sup>. However, in a recent systematic review and meta-analysis, they were able to determine that there is no significant association between CagA and the development of MALT lymphoma (extranodal marginal zone B cell lymphoma) (OR: 1.30; 95% CI: 0.906-1.866) and an inverse association between VacA and the risk of gastric MALT lymphoma (OR: 0.92, 95% CI: 0.57-1.50)<sup>(59)</sup>; Interestingly, CagA translocated into B cells plays a crucial role in the development of DLBCL, and a significant association was observed with this type of lymphoma (OR: 6.43; 95% CI: 2.45-16-84)<sup>(59)</sup>. The role of bacterial genotypes in the pathogenesis of FG remains unclear. This study demonstrated an association between FG and *H. pylori* infection (OR: 13.41, 95% CI: 1.7-103,  $p=0.01$ ), and the iceA1 genotype was more frequent in FG<sup>(99)</sup>.

On the other hand, it is possible that the epigenetic mechanisms may contribute to transforming FG into malignancy<sup>(100)</sup>. MicroRNAs (miRNAs) are non-coding RNAs that can bind to messenger RNA (mRNA) molecules, induce RNA degradation, and lead to post-transcriptional gene silencing<sup>(101)</sup>. miR-150 can modulate the c-Myb transcription factor and influence B cell differentiation<sup>(102)</sup>, and its capacity as a tumor suppressor in DLBCL<sup>(103)</sup>. miR-203 has also been linked to the transformation of gastritis to MALT lymphoma due to promoter methylation that leads to the deregulation of ABL1<sup>(104)</sup>. ABL1 serves as a receptor in B cells, signaling through direct interaction with the BCR and the co-receptor CD19<sup>(105)</sup>. Overexpression of ABL1 has been associated with hematopoietic malignancies such as chronic lymphocytic leukemia<sup>(106)</sup>. ABL1 overexpression has been linked to constitutively active BCR signaling and NF- $\kappa$ B activation<sup>(106)</sup>; that is, this signaling pathway contributes crucially to the genesis of MALT<sup>(64)</sup>.

For another part, the role played by the activation of cytidine deaminase (AID) has been raised, some studies have clarified its role in developing low-grade MALT lymphomas because AID is necessary for developing germinal centers<sup>(107,108)</sup>. AID is recognized as the enzyme responsible for regulating the immunoglobulin gene to initiate class switch recombination (CSR), resulting in immunological diversity and chromosomal translocation of the c-MYC transcription factor<sup>(109,110)</sup>. Unfortunately, AID's

function as a genome mutator may target the generation of somatic mutations in several host genes from non-lymphoid and lymphoid tissues, contributing to tumorigenesis<sup>(110-112)</sup>. In particular, aberrant AID expression can be triggered by several pathogenic factors, including *H. pylori* infection and stimulation of proinflammatory cytokines, while AID expression is absent under physiological conditions<sup>(111,112)</sup>. Therefore, aberrant AID activity in epithelial tissues may provide the critical link between inflammation, somatic mutations, and cancer development<sup>(111-113)</sup> (see Figure 3). *H. pylori* infection increases AID expression through the NF- $\kappa$ B pathway in gastric cells, and with the accumulation of the p53 mutation<sup>(111)</sup>. Likewise, AID activity contributes to lymphomagenesis through the aberrant somatic hypermutation of five other circumscribed proto-oncogenes, such as PIM1, PAX5, RhoH/TTF, and c-MYC<sup>(114-118)</sup>.

Additionally, some genetic abnormalities associated with MALT lymphomas have been identified, the most common being the t(11;18), (q21;q21) translocation, which generates a chimeric protein product of the API-MALT2 fusion that can increase inhibition of apoptosis conferring more remarkable survival to MALT lymphomas. This translocation should be considered after treatment failure or remission following *H. pylori* eradication<sup>(119)</sup>. Other alterations have been described such as the translocation (p22;q32), t(14;18), (q32;q21), t(3;14), (p14.1;q32)<sup>(87)</sup>. Similarly, there is a high prevalence of HLA-DQA1\*0103, HLADQB1\*0601 and the R702W mutation in the NOD2/CARD15 gene<sup>(120,121)</sup>. Additionally, the presence of TNF-857T has been linked<sup>(122)</sup>, and one TLR type 4 allele (TLR4 Asp299Gly)<sup>(87)</sup>.

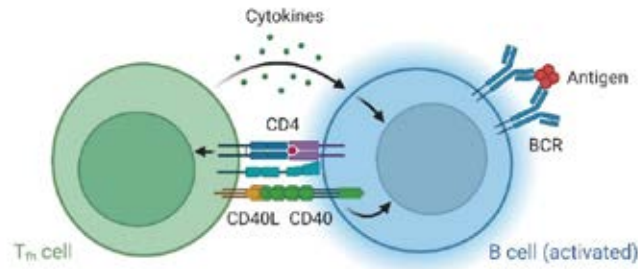
Similarly, at the pathological level, mutations of the p53 protein and the BCL-10 gene are described, which are acquired by the capacity for constitutive activation of NF- $\kappa$ B signal pathway independent of antigenic stimulation when they are overexpressed through the control of promoter regions or hyperactive enhancers from chromosomal translocations<sup>(52,70,81,123-128)</sup>. The BCL-10 gene, located on chromosome 1, is regulated by the IgH immunoglobulin gene on chromosome 14 and results in anarchic expression of the BCL-10 gene<sup>(129)</sup>. In that order, the expression of the BCL-10 protein, which is found at the intracellular level, is essential for the development, differentiation, and function of mature B and T cells<sup>(87)</sup>. Likewise, BCL-10 (B-cell lymphoma 10) protein may be involved in resistance to antimicrobial therapy. A study conducted by Yepes *et al.*<sup>(124)</sup>. In a study of a Colombian population, the significance of t(11;18)(q21;q21), BCL-10 expression, and *H. pylori* infection in MALT lymphoma was evaluated. The study found that MALT lymphoma cases positive for the translocation and those with nuclear BCL-10 overexpression showed a 66% eradication rate of *H. pylori*.

## Diagnosics

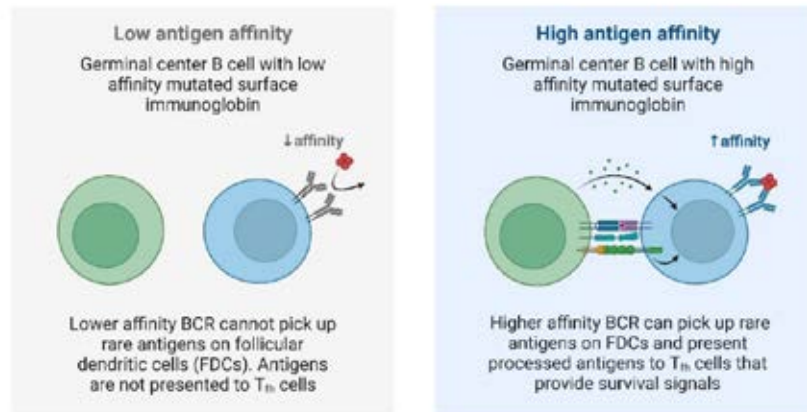
FG diagnosis can be made through endoscopic study and histopathological analysis, considered the standard diagnostic methods to confirm the pathology<sup>(23,130)</sup>. During upper endoscopy, gastric mucosa with multiple nodular formations of a uniform spectrum, predominantly in the

## Somatic Hypermutation in Germinal Center B-cells

### 1. B cell activation



### 2. Somatic hypermutation of immunoglobulin V regions in rapidly proliferating germinal center B cells

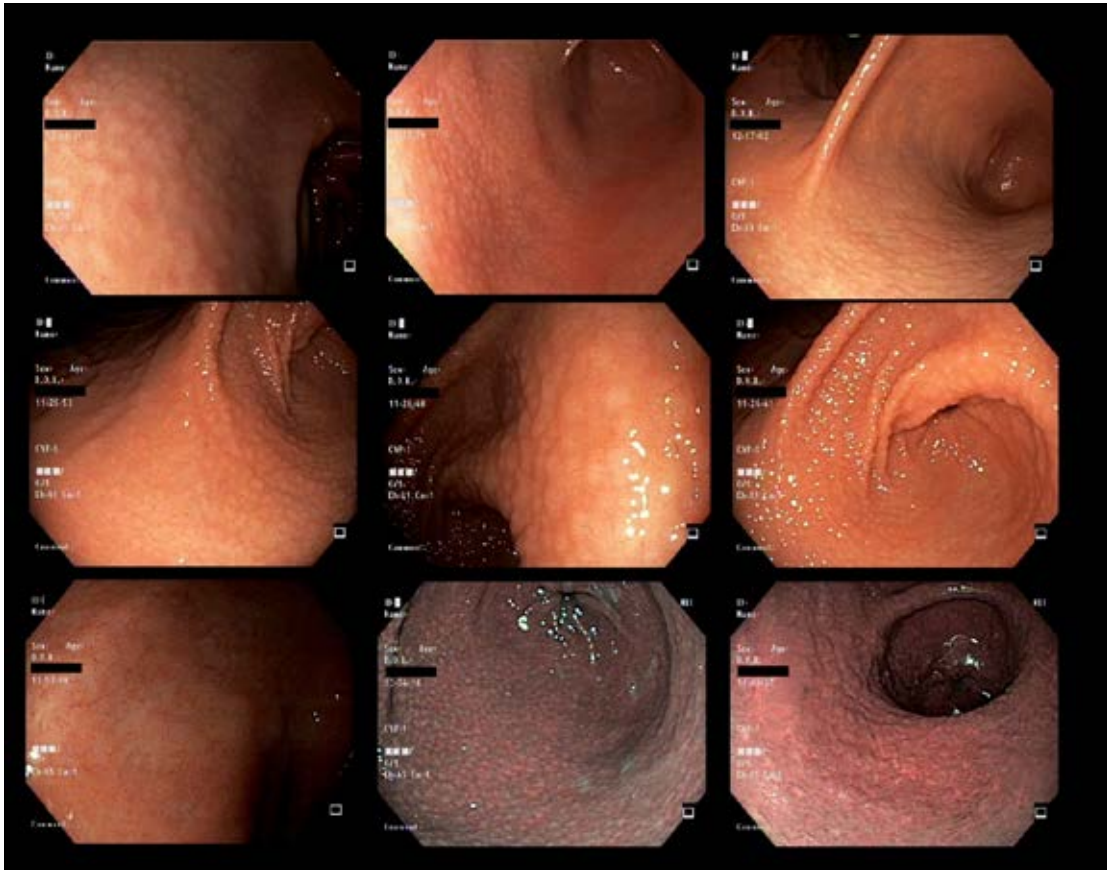


**Figure 3.** Somatic hypermutation and class recombination for its activation by activated cytidine deaminase (AID) in lymphoid proliferation. In the first phase, the activation of B cells through the interaction of CD40-CD40L and MHC-TCR allows the release of cytokines. The B cell, when active, recognizes the antigens of the foreign agent through the BCR. In the second phase, somatic hypermutation of the V regions of the immunoglobulin in the proliferation of the germinal centers of B cells, which the germinal centers are divided by a low and high antigen affinity of the immunoglobulin of the mutated surface for the antigen, which leads to the BCR can pick up rare antigens on follicular dendritic cells (FDC) and present processed antigens to Th cells, that provides survival signals. Source: The authors.

antral location, is observed. However, it can extend to other regions of the gastric body, such as small elevations with superficial erosions and, eventually, ulcerations<sup>(23,131,132)</sup> (see Figure 4). These findings have been documented in both adults and the pediatric population<sup>(133,134)</sup>. Although FG is a designation that is not found in the Sydney classification system for gastritis<sup>(23,135)</sup>. In the Kyoto classification, if the term "nodularity" has been added, which is described with the findings in *H. pylori* infection<sup>(6,131,136,137)</sup>. It must also be considered that FG and MALT lymphoma may overlap and cause difficult endoscopic diagnosis, mainly when FG is present in the body<sup>(31,131,138)</sup>. Similarly, the histopathological obstacle in differentiating FG and MALT lymphoma has been described<sup>(139-142)</sup>. According to this, Hummel *et al.*<sup>(141)</sup>, they investigated B cell clonality in cases of chronic

gastritis (Wotherspoon scores 1 and 2, N=53), gastric MALT lymphoma (Wotherspoon score 5, N=26), and ambiguous histology (Wotherspoon scores 3 and 4, N= 18). The authors noted that B cell clonality was found in 1/53 cases of chronic gastritis (1.9%), 24/26 cases of lymphoma (92.3%), and 4/18 cases with ambiguous histology (22.2%). These similarities and overlaps in pathological, cellular, and molecular features between FG and MALT lymphoma entities are interesting for further research<sup>(131,141)</sup>.

The confirmative diagnosis of the disease depends on the histopathological study of the gastric mucosa in which a mixed inflammatory aggregate and lymphoid follicles with a germinal center are recognized. It is expected to find PMN cells such as neutrophils, ulceration, erosion, and extensive



**Figure 4.** Endoscopic findings of FG.

Source: Provided by the authors Arnoldo Riquelme and Gonzalo Latorre at Pontificia Universidad Catolica de Chile.

fibrosis<sup>(143,144)</sup>. The histological findings of a low-grade MALT lymphoma are expressed with cell proliferation in the presence of the centrocyte, cellular infiltration of plasma, and lymphoepithelial lesions defined as an invasion and partial destruction of the gastric glands and crypts, which are the main characteristics of lymphoma<sup>(145,146)</sup>. However, differentiating between low-grade MALT lymphoma and FG sometimes represents a real diagnostic challenge. Furthermore, immunohistochemistry, cytogenetics, and molecular biology studies are helpful in these cases<sup>(147)</sup>. Furthermore, the analysis of aberrant genes for heavy chain immunoglobulins and polymerase chain reaction (PCR) is widely used as a complementary method<sup>(141)</sup>.

It is worth mentioning that lymphomas exhibit microsatellite instability, allelic imbalance, and trisomy, mainly from chromosome 3, which has been related to the expression of BCL6, FOXP1, and CCR4 in the development<sup>(87,114,148,149)</sup>. Likewise, this abnormality can be determined by immunofluorescence in situ hybridization (FISH) in paraffin blocks<sup>(150)</sup>. Approximately 50% of MALT lymphoma have the chimeric fusion protein API2-MALT1, a product of the t(11:18) translocation that allows constitutive activation of the NF- $\kappa$ B signal pathway; however, this alteration is absent in FG<sup>(83,151)</sup>.

Immunohistochemical studies allow determining the nature of the lymphoid infiltrate and the origin of the MALT lymphoma. However, it is not possible at some point with the tests to distinguish with certainty between a malignant process and a reactive condition. Besides, the combination of the panel of B antigens such as CD3, CD19, CD20, CD79a, and anti-pan cytokeratins has been recommended, in addition to the absence of CD5, CD10, CD23, and the expression of cyclin D1, they are helpful in the detection of infiltration of the mucosal epithelium by B lymphocytes, a condition that defines the lymphoepithelial lesion and establishes the diagnosis of MALT lymphoma<sup>(147,152)</sup>. The use of immunohistochemical markers to determine the cellular monoclonality of neoplasms has a limited contribution, given that this finding is also shared with some lymphoid follicles of FG<sup>(125)</sup>. It has been shown that in this pathology, cells can show variable proliferation, such as polyclonal, monoclonal, and oligoclonal<sup>(26)</sup>. From what was mentioned above, pseudolymphoma, the name of this entity, is derived. For the reasons previously stated, the Committee for the Classification of Tumors of the Digestive System of the WHO establishes the histopathological study as the gold standard for the differentiation between reactive and neoplastic infiltrate, while additional studies



may represent valuable tools in some instances<sup>(153)</sup>. The findings that favor the diagnosis of pseudolymphoma in the biopsy are the proliferation of blood vessels, lymphoid follicles with evident germinal centers, and the absence of lymphoepithelial lesions. A grading system for lesions has been created from 0 to 5 to provide greater security in the diagnosis, with grade 0 being normal and stage 5 defining low-grade lymphoma. This system includes parameters such as lymphoid aggregates, secondary lymphoid follicles, immature lymphocytes, and infiltration of the epithelium<sup>(140,154)</sup>.

## Treatment

*H. pylori* eradication is the standard initial treatment for MALT lymphomas in all stages. However, 70-80% of the pathological condition in stage I is remitted long-term<sup>(1,155,156)</sup>. On the other hand, in patients who are negative for *H. pylori* after routine diagnostics exclude infection, the cure is obtained in 30% of patients, so it should always be considered a first step in management<sup>(1,157)</sup>. However, in MALT lymphomas that transform into DLBCL, in some cases, the same benefit is obtained, mainly when they are dependent on positive *H. pylori*<sup>(158)</sup>. Unlike DLBCL, which lacks histological evidence of MALT lymphoma or negative-*H. pylori*, it has been proven that they are entities with different biological and molecular behaviors<sup>(159)</sup>. Insofar as DLBCL that do not respond to *H. pylori* eradication, chemotherapy remains the standard therapeutic option<sup>(158)</sup>. One study found that patients with MALT lymphoma negative for *H. pylori* had a more advanced clinical stage than patients with positive-*H. pylori* ( $p=0.023$ ). The t(11;18)/API2-MALT1 frequency did not differ between positive-*H. pylori* cases (45.5%) and negative-*H. pylori* cases (55.6%) and only 38/51 (74.5%) positive-*H. pylori* cases achieved complete regression after eradication, while 40% were negative-*H. pylori* cases achieved regression, which allows us to infer that the evaluation of t(11;18)/API2-MALT1 should be considered after failure of remission due to *H. pylori* eradication<sup>(119)</sup>.

Although more studies are required in this regard, recent literature indicates that *H. pylori* eradication is helpful in reversing FG with the usual treatments<sup>(154,160)</sup>. In a clinical trial on 66 patients with FG associated with *H. pylori* (31 with FG vs 35 with NFG), triple therapy for *H. pylori* eradication was administered. At two weeks of follow-up, 44 patients completed treatment (21 with FG vs. 23 with NFG), of which the *H. pylori* eradication in the FG group was 43% and in the NFG group, it was 74%<sup>(154)</sup>. Furthermore, lymphoid follicles in 43% of FG patients disappeared after eradication therapy. The above highlights the importance of corroborating bacterial eradication after treatment<sup>(154)</sup>.

## Conclusions

FG is a form of chronic gastritis characterized by a dense mixed inflammatory infiltrate with the formation of secondary lymphoid follicles in the lamina propria. The incidence in Colombia is 8.4%, and its etiopathogenesis is

influenced by factors such as age, gender, immune response, and *H. pylori* infection.

The diagnosis is established by upper digestive endoscopy and careful histopathological analysis of the gastric mucosa. The most common site is the antrum, so taking biopsies by the gastroenterologist must include representative sampling at this level.

The disease's importance derives from *H. pylori*'s fundamental role in the FG. On the other hand, the bacterial load is not matched with cytokines levels in the inflammatory response, which suggests an alternative mechanism of immune evasion. Further research is crucial for resolving these enigmatic questions.

Treatment for *H. pylori* eradication is the backbone of managing FG, and its elimination must be verified for UBT (urea breath test), RUT (rapid urea test), or stool antigen tests according to available resources and geographic differences.

The field of research is broad and requires knowledge of the entity. In this sense, cellular and molecular mechanisms studies of *H. pylori* and FG are necessary. Furthermore, prospective studies must be carried out to demonstrate the association between FG and progression to MALT lymphoma.

## REFERENCES

1. Malfertheiner P, Camargo MC, El-Omar E, Liou JM, Peek R, Schulz C, et al. Helicobacter pylori infection. Nat Rev Dis Primers. 2023;9(1):19. doi: 10.1038/s41572-023-00431-8.
2. Malfertheiner P, Link A, Selgrad M. Helicobacter pylori: perspectives and time trends. Nat Rev Gastroenterol Hepatol. 2014;11:628-638. doi: 10.1038/nrgastro.2014.99.
3. Hooi JKY, Lai WY, Ng WK, Suen MMY, Underwood FE, Tanyingoh D, et al. Global prevalence of Helicobacter pylori infection: systematic review and meta-analysis. Gastroenterology. 2017;153:420-429.
4. Tang Y, Tang G, Pan L, Zhu H, Zhou S, Wei Z. Clinical factors associated with initial Helicobacter pylori eradication therapy: a retrospective study in China. Scientific Reports. 2020;10:15403. doi: 10.1038/s41598-020-72400-0.
5. Eusebi LH, Zagari RM, Bazzoli F. Epidemiology of Helicobacter pylori infection. Helicobacter. 2014;19(Suppl 1):1-5.
6. Sugano K, Tack J, Kuipers EJ, Graham DY, El-Omar EM, Miura S, et al. Kyoto global consensus report on Helicobacter pylori gastritis. Gut. 2015;64:1353-1367.
7. Yamaoka Y. How to eliminate gastric cancer-related death worldwide? Nat Rev Clin Oncol. 2018;15:407-408.
8. Tshibangu-Kabamba E, Yamaoka Y. Helicobacter pylori infection and antibiotic resistance — from biology to clinical implications. Nat Rev Gastroenterol Hepatol. 2021;18:613-629. doi: 10.1038/s41575-021-00449-x.
9. Humans IWGotEoCRt. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Red Meat and Processed Meat. Lyon (FR): International Agency for Research on Cancer; 2018.
10. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Lyon, 7-14 June 1994. Schistosomes, liver flukes and Helicobacter pylori. IARC Monogr Eval Carcinog Risks Hum. 1994;61:1-241.

11. World Health Organization. ICD-11 Revision [Internet]. Ginebra: WHO; 2019. Disponible en: <http://www.who.int/classifications/icd/revision/en/>
12. Malfertheiner P, Megraud F, Rokkas T, Gisbert JP, Liou JM, Schulz C, et al. Management of Helicobacter pylori infection: the Maastricht VI/Florence consensus report. Gut. 2022;gutjnl-2022-327745.
13. Mejia CR, Vera CA, Huiza-Espinoza L. Association between follicular gastritis and Helicobacter pylori in children seen at a public hospital in Peru. Rev Gastroenterol Mex. 2016;81(2):80-5.
14. Ma Z-Q TT, Nihei Z, Sugihara K, Nakamura K. Follicular gastritis associated with Helicobacter pylori. J Med Dent Sci. 2001;1:39-47.
15. Miyamoto M, Haruma K, Hiyama T, Kamada T, Masuda H, Shimamoto F, et al. High incidence of B-cell monoclonality in follicular gastritis: a possible association between follicular gastritis and MALT lymphoma. Virchows Arch. 2002;440(4):376-80.
16. Sagaert X, Van Cutsem E, De Hertogh G, Geboes K, Tousseyn T. Gastric MALT lymphoma: a model of chronic inflammation-induced tumor development. Nat Rev Gastroenterol Hepatol. 2010;7(6):336-346. doi: 10.1038/nrgastro.2010.58
17. Bujanover Y, Konikoff F, Baratz M. Nodular gastritis and Helicobacter pylori. J Pediatr Gastroenterol Nutr. 1990;11:41-44.
18. Bahú Mda G, da Silveira TR, Maguilnick I, Ulbrich-Kulczynski J. Endoscopic nodular gastritis: an endoscopic indicator of high-grade bacterial colonization and severe gastritis in children with Helicobacter pylori. J Pediatr Gastroenterol Nutr. 2003;36(2):217-222.
19. Carlson SJ, Yokoo H, Vanagunas A. Progression of gastritis to monoclonal B-cell lymphoma with resolution and recurrence following eradication of Helicobacter pylori. JAMA. 1996;275(12):937-9.
20. Asano N, Iijima K, Koike T, Imatani A, Shimosegawa T. Helicobacter pylori-negative gastric mucosa-associated lymphoid tissue lymphomas: A review. World J Gastroenterol. 2015;21(26):8014-20.
21. De Giacomo FR, Villani L, Lisato L, Diegoli N, Donnadini A, et al. Helicobacter pylori infection and chronic gastritis: clinical, serological and histologic correlations in children treated with amoxicillin and colloidal bismuto subcitrate. J Pediatr Gastroenterol Nutr. 1991;11: 310-316.
22. Raymond J, Bergeret MBP, Mensah K, Dupont C. A 2-year study of Helicobacter pylori in children. J Clin Microbiol. 1994;32:461-3.
23. Romero-Flores JL, Fernandez-Rivero JA, Marroquín-Fabian E, Téllez-Ávila FI, Sánchez-Jiménez BA, Juárez-Hernández E, et al. Diagnostic accuracy of nodular gastritis for H. pylori infection. Ther Clin Risk Manag. 2017;13:9-14.
24. Kalali B, Formichella L, Gerhard M. Diagnosis of Helicobacter pylori: Changes towards the Future. Diseases. 2015;3(3):122-35.
25. Chey WD, Leontiadis GI, Howden CW, Moss SF. ACG Clinical Guideline: Treatment of Helicobacter pylori Infection. Am J Gastroenterol. 2017;112(2):212-39.
26. Lo WY LJ, Chan YK, Lai LS, Yeung YW, Lo ST, Tsui WM, Ng CS. Instability of clonality in gastric lymphoid infiltrates: a study with emphasis on serial biopsies. Am J Surg Pathol. 2005;29:1582-92.
27. Hong K, Tae Wong N, Seuong -Yon B. Nodular Gastritis A Pathologic Findings In Children And Young Adults With Helicobacter Pylori Infection. Yonsei Med J. 2007;48(2):240-246.
28. Wyatt JI, Rathbone BJ. Immune response of the gastric mucosa to Campylobacter pylori. Scand J Gastroenterol. 1988;142:44-9.
29. Eidt S, Stolte M. Prevalence of lymphoid follicles and aggregates in Helicobacter pylori gastritis in antral and body mucosa. J Clin Pathol. 1993;46:832-5.
30. Genta RM, Hammer HW. The significance of lymphoid follicles in the interpretation of gastric biopsy specimens. Arch Pathol Lab Med. 1994;118:740-3.
31. Chen XY, Liu WZ, Shi Y, Zhang DZ, Xiao SD, Tytgat GN. Helicobacter pylori associated gastric diseases and lymphoid tissue hyperplasia in gastric antral mucosa. J Clin Pathol. 2002;55(2):133-7. doi: 10.1136/jcp.55.2.133
32. Chen MJ, Wang TE, Chang WH, Liao TC, Lin CC, Shih SC. Nodular gastritis: an endoscopic indicator of Helicobacter Pylori infection. Dig Dis Sci. 2007;52(10):2662-6.
33. Shimatani T, Inoue M, Iwamoto K, Hyogo H, Yokozaki M, Saeki T, et al. Gastric acidity in patients with follicular gastritis is significantly reduced, but can be normalized after eradication for Helicobacter pylori. Helicobacter. 2005;10(3):256-65.
34. Choi HJ, Lee SY, Lee JH, Seol DC, Kim SY, Choi HJ, et al. Two Atypical Cases of Nodular Gastritis: A Poorly Differentiated Gastric Adenocarcinoma and a Pseudo-Low Grade Gastric MALT Lymphoma. Gastroenterology Res. 2010;3(1):41-5.
35. Martínez-Marín JD, Henao-Riveros SC. Hiperplasia linfocítica gástrica e infección por Helicobacter pylori en adultos colombianos. Rev Col Gastroenterol. 2009;24(2): 148-156.
36. Melo-Peñalosa MA, Mendoza-Rodríguez A. Frequency of morphological changes in gastric biopsies associated with Helicobacter pylori infection. Acta Med Colomb. 2021;46(3):25-31. doi: 10.36104/amc.2021.1987.
37. Zaitoun AM. The prevalence of lymphoid follicles in Helicobacter pylori associated gastritis in patients with ulcers and non-ulcer dyspepsia. J Clin Pathol. 1995;48:325-9.
38. Zerbib F, Vialette G, Cayla R, Rudelli A, Sauvet P, Bechade D, Seurat PL, Lamouliatte H. Follicular gastritis in adults. Relations with Helicobacter pylori, histological and endoscopic aspects. Gastroenterol Clin Biol. 1993;17(8-9):529-34.
39. Bedoya A, Arcos M, Sansón F, del Castillo G. Helicobacter pylori y cambios histológicos de la mucosa gástrica en menores de diez años. Pasto, 1999. Rev Colomb Gastroenterol. 2002;17: 36-42.
40. Martínez JD, Henao SC, Granados C. La Gastritis crónica atrófica y la edad. Rev Colomb Gastroenterol. 2007;22: 17-22.
41. Ladas S, Rokkas T, Georgopoulos S, Kitsanta P, Liatsos C, Eustathiadou P, et al. Predictive factors and prevalence of follicular gastritis in adults with peptic ulcer and nonulcer dyspepsia. Dig Dis Sci. 1999;44(6):1156-60.
42. Rafeey M, Jafari Rouhi AH, Gassemi BA, Rouhi AJ. Relationship between endoscopic nodular gastritis and Helicobacter pylori infection in children. Indian J Gastroenterol. 2004;23(4):138-139.
43. Mansilla-Vivar R, Serrano CA, Palma C, Vera M, Hernandez C, Pizarro M, et al. High Helicobacter pylori Bacterial Load and Low Cytokine Expression Levels Are Associated with Nodular Gastropathy. Dig Dis Sci. 2020;65(2):565-75.
44. Marcellis L, Tousseyn T, Sagaert X. MALT Lymphoma as a Model of Chronic Inflammation-Induced Gastric Tumor Development. Curr Top Microbiol Immunol. 2019;421:77-106. doi: 10.1007/978-3-030-15138-6\_4
45. Isaacson PG. Update on MALT lymphomas. Best Pract Res Clin Haematol [Internet]. 2005;18(1):57-68. doi: 10.1016/j.beha.2004.08.003.
46. Bardhan PK. Epidemiological features of Helicobacter pylori infection in developing countries. Clin Infect Dis. 1997;25(5):973-978. doi: 10.1086/516067.
47. Asenjo LM, Gisbert JP. [Prevalence of Helicobacter pylori infection in gastric MALT lymphoma: a systematic review]. Rev Esp Enferm Dig. 2007;99(7):398-404.
48. Garhart C, Czinn S. Helicobacter pylori infection: Review of pathogenesis and immunity. Int Semin Paediatr Gastroenterol Nutr. 2004;12:3-7.

49. Bergman MP, D'Ellos MM. Cytotoxic T cells in H. pylori-related gastric autoimmunity and gastric lymphoma. *J Biomed Biotechnol.* 2010;2010:104918. doi: 10.1155/2010/104918.
50. Yamaoka Y. Mechanisms of disease: Helicobacter pylori virulence factors. *Nat Rev Gastroenterol Hepatol.* 2010;7:629-641. doi: 10.1038/nrgastro.2010.154.
51. Malfertheiner P, Megraud F, O'Morain CA, Gisbert JP, Kuipers EJ, Axon AT, et al. Management of Helicobacter pylori infection-the Maastricht V/Florence Consensus Report. *Gut.* 2017;71:1724-1762. doi: 10.1136/gutjnl-2016-312288.
52. Araújo GRL, Marques HS, Santos MLC, da Silva FAF, da Brito BB, Correa Santos GL, et al. Helicobacter pylori infection: How does age influence the inflammatory pattern? *World J Gastroenterol.* 2022;28(4):402-11.
53. Onal IK, Gokcan H, Benzer E, Bilir G, Oztas E. What is the impact of Helicobacter pylori density on the success of eradication therapy: a clinico-histopathological study. *Clin Res Hepatol Gastroenterol.* 2013;37(6):642-6.
54. Elitsur Y, Lawrence Z, Triest WE. Distribution of Helicobacter pylori organisms in the stomachs of children with H. pylori infection. *Hum Pathol.* 2002;33(11):1133-5.
55. Camorlinga-Ponce M, Aviles-Jimenez F, Cabrera L, Hernández-Pando R, Muñoz O, Soza J, et al. Intensity of inflammation, density of colonization and interleukin-8 response in the gastric mucosa of children infected with Helicobacter pylori. *Helicobacter.* 2003;8(5):554-60.
56. Xu XQ, Wang ZH, Liao JX, Chen XY, Liu WZ, Xiao SD, et al. Predictive value of neutrophil infiltration as a marker of Helicobacter pylori infection. *World J Gastroenterol.* 2012;18(36):5101-5.
57. Hatakeyama M, Higashi H. Helicobacter pylori CagA: a new paradigm for bacterial carcinogenesis. *Cancer Sci.* 2005;96(12):835-843. doi: 10.1111/j.1349-7006.2005.00130.x.
58. Li SP, Chen XJ, Sun AH, Zhao JF, Yan J. CagA(+) H. pylori induces Akt1 phosphorylation and inhibits transcription of p21(WAF1/CIP1) and p27(KIP1) via PI3K/Akt1 pathway. *Biomed Environ Sci.* 2010;23(4):273-278. doi: 10.1016/S0895-3988(10)60063-3.
59. Keikha M, Sahebkar A, Yamaoka Y, Karbalaeei M. Helicobacter pylori cagA status and gastric mucosa-associated lymphoid tissue lymphoma: a systematic review and meta-analysis. *J Health Popul Nutr.* 2022;41:2.
60. Yong X, Tang B, Li BS, Xie R, Hu CJ, Luo G, et al. Helicobacter pylori virulence factor CagA promotes tumorigenesis of gastric cancer via multiple signaling pathways. *Cell Commun Signal.* 2015;13:30.
61. Machado AMD, Figueiredo C, Touati E, Máximo V, Sousa S, Michel V, et al. Helicobacter pylori infection induces genetic instability of nuclear and mitochondrial DNA in gastric cells. *Clin Cancer Res.* 2009;15(9):2995-3002.
62. Fox JG, Wang TC. Inflammation, atrophy, and gastric cancer. *J Clin Invest.* 2007;117(1):60-9.
63. Jaffe ES, Harris NL, Stein H, Isaacson PG. Classification of lymphoid neoplasms: the microscope as a tool for disease discovery. *Blood.* 2008;112(12):4384-99.
64. Lim KH, Yang Y, Staudt LM. Pathogenetic importance and therapeutic implications of NF- $\kappa$ B in lymphoid malignancies. *Immunol Rev.* 2012;246(1):359-78.
65. Lin WC, Tsai HF, Kuo SH, Wu MS, Lin CW, Hsu PI, et al. Translocation of Helicobacter pylori CagA into Human B lymphocytes, the origin of mucosa-associated lymphoid tissue lymphoma. *Cancer Res.* 2010;70(14):5740-8.
66. Umehara S, Higashi H, Ohnishi N, Asaka M, Hatakeyama M. Effects of Helicobacter pylori CagA protein on the growth and survival of B lymphocytes, the origin of MALT lymphoma. *Oncogene.* 2003;22(51):8337-42.
67. Krisch LM, Posselt G, Hammerl P, Wessler S. CagA Phosphorylation in Helicobacter pylori-Infected B Cells Is Mediated by the Nonreceptor Tyrosine Kinases of the Src and Abl Families. *Infect Immun.* 2016;84(9):2671-80.
68. Bagheri V, Memar B, Momtazi AA, Sahebkar A, Gholamin M, Abbaszadegan MR. Cytokine networks and their association with Helicobacter pylori infection in gastric carcinoma. *J Cell Physiol.* 2018;233(4):2791-803.
69. Touati E, Michel V, Thiberge JM, Wuscher N, Huerre M, Labigne A. Chronic Helicobacter pylori infections induce gastric mutations in mice. *Gastroenterology.* 2003;124(5):1408-19.
70. Traci L, Testerman JM. Beyond the stomach: An updated view of Helicobacter pylori pathogenesis, diagnosis, and treatment. *World J Gastroenterol.* 2014;28(36):12781-808.
71. Backert S, Clyne M. Pathogenesis of Helicobacter pylori infection. *Helicobacter.* 2011;16 Suppl 1:19-25.
72. Bagheri N, Salimzadeh L, Shirzad H. The role of T helper 1-cell response in Helicobacter pylori-infection. *Microb Pathog.* 2018;123:1-8.
73. Imlay JA. Cellular defenses against superoxide and hydrogen peroxide. *Annu Rev Biochem.* 2008;77:755-76.
74. Spiro S, D'Autréaux B. Non-heme iron sensors of reactive oxygen and nitrogen species. *Antioxid Redox Signal.* 2012;17(9):1264-76.
75. Vázquez-Torres A. Redox active thiol sensors of oxidative and nitrosative stress. *Antioxid Redox Signal.* 2012;17(9):1201-14.
76. Iseki K, Tatsuta M, Iishi H, Baba M, Mikuni T, Hirasawa R, et al. Attenuation by methionine of monochloramine-enhanced gastric carcinogenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine in Wistar rats. *Int J Cancer.* 1998;76(1):73-6.
77. Iishi H, Tatsuta M, Baba M, Mikuni T, Yamamoto R, Iseki K, et al. Enhancement by monochloramine of the development of gastric cancers in rats: A possible mechanism of Helicobacter pylori-associated gastric carcinogenesis. *J Gastroenterol.* 1997;32:435-441. doi: 10.1007/BF02934080.
78. Romagnani S, Parronchi P, D'Ellos MM, Romagnani P, Annunziato F, Piccinni MP, et al. An update on human Th1 and Th2 cells. *Int Arch Allergy Immunol.* 1997;113:153-156.
79. Craig VJ, Cogliatti SB, Arnold I, Gerke C, Balandat JE, Wündisch T, et al. B-cell receptor signaling and CD40 ligand-independent T cell help cooperate in Helicobacter-induced MALT lymphomagenesis. *Leukemia.* 2010;24(6):1186-96.
80. García M, Bellosillo B, Sánchez-González B, García-Payarols F, Seoane A, Ferrer AM, et al. Study of regulatory T-cells in patients with gastric malt lymphoma: influence on treatment response and outcome. *PLoS One.* 2012;7(12):e51681.
81. Uhl B, Prochazka KT, Fechter K, Pansy K, Greinix HT, Neumeister P, et al. Impact of the microenvironment on the pathogenesis of mucosa-associated lymphoid tissue lymphomas. *World J Gastrointest Oncol.* 2022;14(1):153-62.
82. D'Ellos MM, Manghetti M, Almerigogna F, Amedei A, Costa F, Burrioni D, et al. Different cytokine profile and antigen-specificity repertoire in Helicobacter pylori-specific T cell clones from the antrum of chronic gastritis patients with or without peptic ulcer. *Eur J Immunol.* 1997;27:1751-1755.
83. Rosebeck S, Madden L, Jin X, Gu S, Apel IJ, Appert A, et al. Cleavage of NIK by the API2-MALT1 fusion oncoprotein leads to noncanonical NF- $\kappa$ B activation. *Science.* 2011;331(6016):468-72.
84. Jardin F. NF- $\kappa$ B Pathway and Hodgkin Lymphoma. *Biomedicines.* 2022;10:2153. doi: 10.3390/biomedicines10092153.
85. Bauer B, Moese S, Bartfeld S, Meyer TF, Selbach M. Analysis of cell type-specific responses mediated by the type IV secretion system of Helicobacter pylori. *Infect Immun.* 2005;73(8):4643-4652.

86. Nishikori M. Classical and alternative NF- $\kappa$ B activation pathways and their roles in lymphoid malignancies. *J Clin Exp Hematopathol.* 2005;45:15-24.
87. Ronchi A, Montella M, Panarese I, Costanzo RMA, Aquino G, De Chiara A, et al. MALT gastric lymphoma: An pathogenetic features. *WCRJ.* 2016;3(2):e715.
88. Elgueta R, Benson MJ, de Vries VC, Wasiuk A, Guo Y, Noelle RJ. Molecular mechanism and function of CD40/CD40L engagement in the immune system. *Immunol Rev.* 2009;229(1):152-72. doi: 10.1111/j.1600-065X.2009.00782.x.
89. Chan JK C, Ng CS, Isaacson PG. Relationship between high-grade lymphoma and low-grade B-cell mucosa-associated lymphoid tissue lymphoma (MALToma) of the stomach. *Am J Pathol.* 1990;136:1153-1164.
90. Villuendas R, Piris MA, Orradre JL, Mollajo M, Rodriguez R, Morente M. Different bcl-2 protein expression in high-grade B-cell lymphomas derived from lymph node or mucosa-associated lymphoid tissue. *Am J Pathol.* 1991;139:989-993.
91. Blosser A, Peru S, Levy M, Marteyn B, Floch P, Sifré E, et al. APRIL-producing eosinophils are involved in gastric MALT lymphomagenesis induced by Helicobacter sp infection. *Sci Rep.* 2020;10(1):14858.
92. Yoon JH, Choi WS, Kim O, Park WS. The role of gastrokine 1 in gastric cancer. *J Gastric Cancer.* 2014;14:147-55.
93. Nardone G, Rippe E, Martin G, Rocco A, Siciliano RA, Fiengo A, et al. Gastrokine 1 expression in patients with and without Helicobacter pylori infection. *Dig Liver Dis.* 2007;39:122-9.
94. Alarcón-Millán, J., Lorenzo-Nazario, S.I., Jiménez-Wences, H. et al. Women with chronic follicular gastritis positive for Helicobacter pylori express lower levels of GKN1. *Gastric Cancer.* 2020;23:754-759. doi: 10.1007/s10120-020-01049-5.
95. Cappell M. S., Garcia A. Gastric and duodenal ulcers during pregnancy. *Gastroenterol Clin North Am.* 1998;27:169-195.
96. Havarad TJ, Sarsfield P, Wotherspoon AC, Steer HW. Increased gastric epithelial cell proliferation in Helicobacter pylori associated follicular gastritis. *J Clin Pathol.* 1996;49(1):68-71.
97. Blaser MJ. Helicobacter pylori and gastric diseases. *BMJ.* 1998;316(7143):1507-10.
98. Kuo SH, Chen LT, Lin CW, Wu MS, Hsu PN, Tsai HJ, et al. Detection of the Helicobacter pylori CagA protein in gastric mucosa-associated lymphoid tissue lymphoma cells: clinical and biological significance. *Blood Cancer J.* 2013;3(7):e125.
99. Carlosama-Rosero YH, Bolaños-Bravo H, Sierra-Tórres CH, Rosero EA. Association of the Helicobacter pylori cagA, vacA, and iceA genotypes with chronic follicular gastritis in a Colombian population at high risk for gastric cancer. *Rev Gastroenterol Mex (Engl Ed).* 2019;84(2):158-64.
100. Thorns C, Kuba J, Bernard V, Senft A, Szymczak S, Feller AC, et al. Deregulation of a distinct set of microRNAs is associated with transformation of gastritis into MALT lymphoma. *Virchows Arch.* 2012;460:371-377. doi: org/10.1007/s00428-012-1215-1.
101. Belair C, Darfeuille F, Staedel C. Helicobacter pylori and gastric cancer: possible role of microRNAs in this intimate relationship. *Clin Microbiol Infect.* 2009;15:806-812. doi: 10.1111/j.1469-0691.2009.02960.x
102. Xiao C, Calado DP, Galler G, Thai TH, Patterson HC, Wang J, et al. MiR-150 controls B cell differentiation by targeting the transcription factor c-Myb. *Cell.* 2007;131:146-159. doi: 10.1016/j.cell.2007.07.021.
103. Roehle A, Hoefig KP, Reipsilber D, Thorns C, Ziepert M, Wesche KO, et al. MicroRNA signatures characterize diffuse large B-cell lymphomas and follicular lymphomas. *Br J Haematol.* 2008;142:732-744. doi: 10.1111/j.1365-2141.2008.07237.x.
104. Craig VJ, Cogliatti SB, Rehauer H, Wündisch T, Müller A. Epigenetic silencing of microRNA-203 dysregulates ABL1 expression and drives Helicobacter-associated gastric lymphomagenesis. *Cancer Res.* 2011;71(10):3616-24.
105. Zipfel PA, Grove M, Blackburn K, Fujimoto M, Tedder TF, Pendergast AM. The c-Abl tyrosine kinase is regulated downstream of the B cell antigen receptor and interacts with CD19. *J Immunol.* 2000;165: 6872-9.
106. Lin K, Glenn MA, Harris RJ, Duckworth AD, Dennett S, Cawley JC, et al. c-Abl expression in chronic lymphocytic leukemia cells: clinical and therapeutic implications. *Cancer Res.* 2006;66:7801-9.
107. Greeve J, Philipsen A, Krause K, Klapper W, Heidorn K, Castle BE, et al. Expression of activation-induced cytidine deaminase in human B-cell non-Hodgkin lymphomas. *Blood.* 2003;101(9):3574-80.
108. Pasqualucci L, Bhagat G, Jankovic M, Compagno M, Smith P, Muramatsu M, et al. AID is required for germinal center-derived lymphomagenesis. *Nat Genet.* 2008;40(1):108-12.
109. Zhang J, Shi Y, Zhao M, Hu H, Huang H. Activation-induced cytidine deaminase overexpression in double-hit lymphoma: potential target for novel anticancer therapy. *Sci Rep.* 2020;10:14164. doi: 10.1038/s41598-020-71058-y.
110. Shikata H, Yakushijin Y, Matsushita N, Sakai A, Sugita A, Nakamura N, et al. Role of activation-induced cytidine deaminase in the progression of follicular lymphoma. *Cancer Sci.* 2012;103(3):415-21.
111. Matsumoto Y, Marusawa H, Kinoshita K, Endo Y, Kou T, Morisawa T, et al. Helicobacter pylori infection triggers aberrant expression of activation-induced cytidine deaminase in gastric epithelium. *Nat Med.* 2007;13(4):470-476. doi: 10.1038/nm1566.
112. Marusawa H, Chiba T. Helicobacter pylori-induced activation-induced cytidine deaminase expression and carcinogenesis. *Curr Opin Immunol.* 2010;22(4): 442-447.
113. Marusawa H. Aberrant AID expression and human cancer development. *Int J Biochem Cell Biol.* 2008;40(8):1399-402.
114. Deutsch AJA, Aigelsreiter A, Staber PB, Beham A, Linkesch W, Guelly C, et al. MALT lymphoma and extranodal diffuse large B-cell lymphoma are targeted by aberrant somatic hypermutation. *Blood.* 2007;109(8):3500-4.
115. Halldórsdóttir AM, Frühwirth M, Deutsch A, Aigelsreiter A, Beham-Schmid C, Agnarsson BA, et al. Quantifying the role of aberrant somatic hypermutation in transformation of follicular lymphoma. *Leuk Res.* 2008;32:1015-1021.
116. Gaidano G, Pasqualucci L, Capello D, Berra E, Deambroggi C, Rossi D, et al. Aberrant somatic hypermutation in multiple subtypes of AIDS-associated non-Hodgkin lymphoma. *Blood.* 2003;102:1833-1841.
117. Dijkman R, Tensen CP, Buettn er M, Niedobitek G, Willemze R, Vermeer MH. Primary cutaneous follicle center lymphoma and primary cutaneous large B-cell lymphoma, leg type, are both targeted by aberrant somatic hypermutation but demonstrate differential expression of AID. *Blood.* 2006;107:4926-4929.
118. Bödör C, Bognár A, Reiniger L, Szepesi A, Tóth E, Kopper L, et al. Aberrant somatic hypermutation and expression of activation-induced cytidine deaminase mRNA in mediastinal large B-cell lymphoma. *Br J Haematol.* 2005;129:373-376.
119. Choi YJ, Kim N, Paik JH, Kim JM, Lee SH, Park YS, et al. Characteristics of Helicobacter pylori-positive and Helicobacter pylori-negative gastric mucosa-associated lymphoid tissue lymphoma and their influence on clinical outcome. *Helicobacter.* 2013;18(3):197-205.

120. Rosenstiel P, Hellmig S, Hampe J, Ott S, Till A, Fischbach W, et al. Influence of polymorphisms in the NOD1/CARD4 and NOD2/CARD15 genes on the clinical outcome of Helicobacter pylori infection. *Cell Microbiol.* 2006;8(7):1188–1198.
121. Zullo A, Hassan C, Cristofari F, Andriani A, De Francesco V, Ierardi E, et al. Effects of Helicobacter pylori Eradication on Early Stage Gastric Mucosa-Associated Lymphoid Tissue Lymphoma. *Clin Gastroenterol Hepatol.* 2010;8(2):105–110.
122. Kuo SH, Yeh KH, Chen LT, Lin CW, Hsu PN, Hsu C, et al. Helicobacter pylori-related diffuse large B-cell lymphoma of the stomach: A distinct entity with lower aggressiveness and higher chemosensitivity. *Blood Cancer J.* 2014;4(6):e220. doi: 10.1038/bcj.2014.40.
123. Mahdavi J, Sondén B, Hurtig M, Olfat FO, Forsberg L, Roche N, et al. Helicobacter pylori SabA adhesin in persistent infection and chronic inflammation. *Science.* 2002;297(5581):573–8.
124. Yepes S, Torres MM, Saavedra C, Andrade R. Gastric mucosa-associated lymphoid tissue lymphomas and Helicobacter pylori infection: A Colombian perspective. *World J Gastroenterol.* 2012;18(7):685–91.
125. Wundiisch T, Neubauer A, Stolte M, Ritter M, Thiede C. B-cell monoclonality is associated with lymphoid follicles in gastritis. *Am J Surg Pathol.* 2003;27:882–887.
126. Yeh KH, Kuo SH, Chen LT, Mao TL, Doong SL, Wu MS, et al. Nuclear expression of BCL10 or nuclear factor kappa B helps predict Helicobacter pylori-independent status of low-grade gastric mucosa-associated lymphoid tissue lymphomas with or without t(11;18)(q21;q21). *Blood.* 2005;106(3):1037–41.
127. Morales-Fuentes GA, Zarate-Osorno A, Quiñónez-Urrego EE, Antonio-Manrique M, Martínez-García CL, Figueroa-Barojas P, et al. p53 expression in the gastric mucosa of patients infected with Helicobacter pylori. *Rev Gastroenterol Mex.* 2013;78(1):12–20.
128. Strehl JD, Hoegel J, Hornicek I, Hartmann A, Riener MO. Immunohistochemical expression of IMP3 and p53 in inflammatory lesions and neoplastic lesions of the gastric mucosa. *Int J Clin Exp Pathol.* 2014;7(5):2091–101.
129. Ruland J, Duncan GS, Elia A, Del Barco Barrantes I, Nguyen L, Plyte S, et al. Bcl-10 is a positive regulator of antigen receptor-induced activation of NF-kappaB and neural tube closure. *Cell.* 2001;104: 33–42.
130. Miyamoto M, Haruma K, Yoshihara M, Hiyama T, Sumioka M, Nishisaka T, et al. Nodular gastritis in adults is caused by Helicobacter pylori infection. *Dig Dis Sci.* 2003;48(5):968–975.
131. Iwamuro M, Tanaka T, Nishida K, Kanzaki H, Kawano S, Kawahara Y, et al. Two cases of gastric mucosa-associated lymphoid tissue (MALT) lymphoma masquerading as follicular gastritis. *Ecancermedalscience.* 2019;13:933.
132. ASGE Standards of Practice Committee, Evans JA, Chandrasekhara V, Chathadi KV, Decker GA, Early DS, et al. The role of endoscopy in the management of premalignant and malignant conditions of the stomach. *Gastrointest Endosc.* 2015;82(1):1–8.
133. Rosh JR, Kurfist LA, Benkov KJ, Toor AH, Bottone EJ, LeLeiko NS. Helicobacter pylori and gastric lymphonodular hyperplasia in children. *Am J Gastroenterol.* 1992;87(1):135–9.
134. Sbei F, Abdullah A, Sullivan S, Merenkov Z. Antral nodularity, gastric lymphoid hyperplasia, and Helicobacter pylori in adults. *J Clin Gastroenterol.* 1996;22(3):227–30.
135. Sipponen P, Price AB. The Sydney System for classification of gastritis 20 years ago. *J Gastroenterol Hepatol.* 2011;26 Suppl 1:31–4.
136. Sugimoto M, Ban H, Ichikawa H, Sahara S, Otsuka T, Inatomi O, et al. Efficacy of the Kyoto Classification of Gastritis in Identifying Patients at High Risk for Gastric Cancer. *Intern Med.* 2017;56(6):579–86.
137. Kamada T, Haruma K, Inoue K, Shiotani A. [Helicobacter pylori infection and endoscopic gastritis -Kyoto classification of gastritis]. *NihonShokakibyogakkaiZasshi.* 2015;112(6):982–93.
138. Lee KS, Yang HR, and Ko JS, Seo JK, Lee HS. A case of gastric MALT lymphoma presenting as nodular gastritis in a child. *Korean J Pediatr Gastroenterol Nutr.* 2008;11(2):187–192.
139. Cheng TI, Tsou MH, and Tsai MP. Early gastric MALT lymphoma. *J Chin Med Assoc.* 2004;67(3):145–148.
140. Wotherspoon Ac, Doglioni C, Isaacson PG. Low-grade gastric B-cell lymphoma of mucosa-associated lymphoid tissue(MALT). A multifocal disease. *Histopathology.* 1992;20:29–34.
141. Hummel M, Oeschger S, Barth TF, Loddenkemper C, Cogliatti SB, Marx A, et al. Wotherspoon criteria combined with B cell clonality analysis by advanced polymerase chain reaction technology discriminates covert gastric marginal zone lymphoma from chronic gastritis. *Gut.* 2006;55(6):782–787. doi: 10.1136/gut.2005.080523.
142. Doglioni C, Ponzone M, Ferreri AJM, Savio A, Gruppo Italiano Patologi Apparato Digerente (GIPAD), Società Italiana di Anatomia Patologica e Citopatologia Diagnostica/International Academy of Pathology, Italian division (SIAPEC/IAP). Gastric lymphoma: the histology report. *Dig Liver Dis.* 2011;43 Suppl 4:S310–318.
143. Perez CA, Dorfman RF. Benign lymphoid hiperplasia of the stomach and duodenum. *Radiology.* 1966;87:505–510.
144. Hyjek E, Kelényi G. Pseudolymphoma of the stomach. A lesion characterized by progressively transformed germinal centers. *Histopathology.* 1982;6:61–68.
145. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, et al. WHO classification of tumours of haematopoietic and lymphoid tissues. IARC Press; 2008.
146. Isaacson PG, Spencer J. Malignant lymphoma of mucosa-associated lymphoid tissue. *Histopathology.* 1987;11(5):445–62.
147. Chan J. K. Gastrointestinal lymphomas: an overview with emphasis on new findings and diagnostic problems. *Semin Diagn Pathol.* 1996;13(4):260–96.
148. Dierlamm J, Wlodarska I, Michaux L. Genetic abnormalities in marginalzoneB-celllymphoma. *HematolOncol.* 2000;18:1–13.
149. Deutsch AJA, Aigelsreiter A, Steinb auer E, Neumeister, P. Distinct signatures of B-cell homeostatic and activation-dependent chemokine receptors in the development and progression of extragastric MALT lymphomas. *J Pathol.* 2008;215:431–444.
150. Blanco R, Lyda M, Davis B, Kraus M, Fenoglio C. Trisomy 3 in gastric lymphomas of extranodal marginal zone B- cell (mucosa-associated lymphoid tissue) origin demonstrated by FISH in intact paraffin tissue sections. *Hum Pathol.* 1990;30:706–711.
151. Baens M, Maes B, Steyls A, Geboes K, Marynen P, De Wolff-Peters C. The product of the t(11-18), an AP12-MLT fusion, marks nearly half of the gastric MALT type lymphomas without large cell proliferation. *An J Pathol.* 2000;156:1433–1439.
152. Pileri SA, Sabattini E. A rational approach to immunohistochemical analysis of malignant lymphomas on paraffin wax sections. *J Clin Pathol.* 1997;50(1):2–4.
153. Wotherspoon A, Chot A, Gascoyne RD, Muller-Hermelinck HK. Lymphoma of the stomach. In Hamilton SR, Aaltonen LA (eds): Pathology and genetics of tumours of the digestive system, World Health Organization classification of tumours. Lyon: IARC Press; 2000. p. 57–61.
154. Mehmet S, Ozdal E, Kamil O, Beşir K, Huseyin D, Nihat A, Cigdem EY, Nusret E. Eradication of Helicobacter pylori in

- follicular and nonfollicular gastritis. *Hepatogastroenterology*. 2009;56(91-92):930-4.
155. Ruskoné-Fourmestraux A, Fischbach W, Aleman BM, Boot H, Du MQ, Megraud F, et al. Gastric extranodal marginal zone B-cell lymphoma of MALT. *Gut*. 2011;60:747-758.
  156. Raderer M, Kiesewetter B, Ferreri AJ. Clinicopathologic characteristics and treatment of marginal zone lymphoma of mucosa-associated lymphoid tissue (MALT lymphoma). *CA Cancer J Clin*. 2016;66:153-171.
  157. Jung K, Kim DH, Seo HI, Gong EJ, Bang CS. Efficacy of eradication therapy in *Helicobacter pylori*-negative gastric mucosa-associated lymphoid tissue lymphoma: A meta-analysis. *Helicobacter*. 2021;26(2):e12774.
  158. Paydas S. *Helicobacter pylori* eradication in gastric diffuse large B cell lymphoma. *World J Gastroenterol*. 2015;21(13):3773-6.
  159. Kuo SH, Cheng AL. *Helicobacter pylori* and mucosa-associated lymphoid tissue: what's new. *Hematology Am Soc Hematol Educ Program*. 2013;2013:109-17.
  160. Dwivedi M, Misra SP, Misra V. Nodular gastritis in adults: clinical features, endoscopic appearance, histopathological features, and response to therapy. *J Gastroenterol Hepatol*. 2008;23(6):943-947.